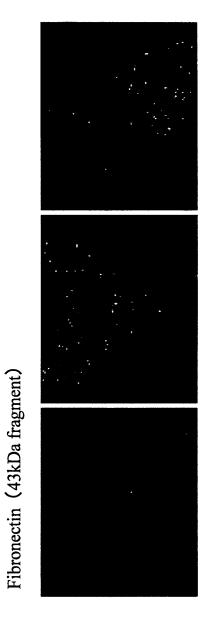
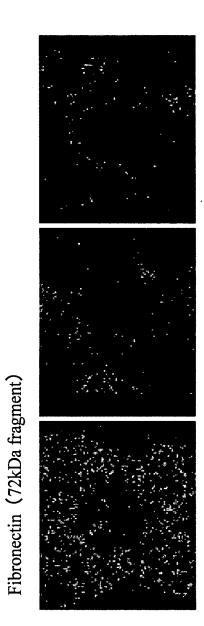
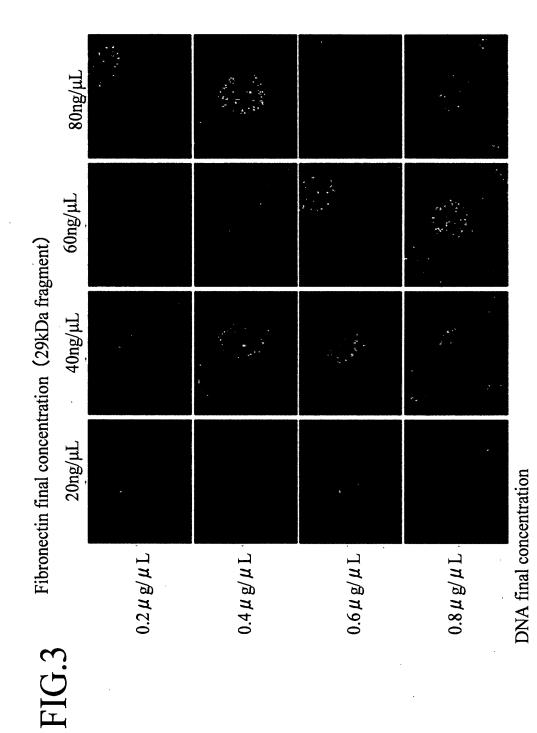
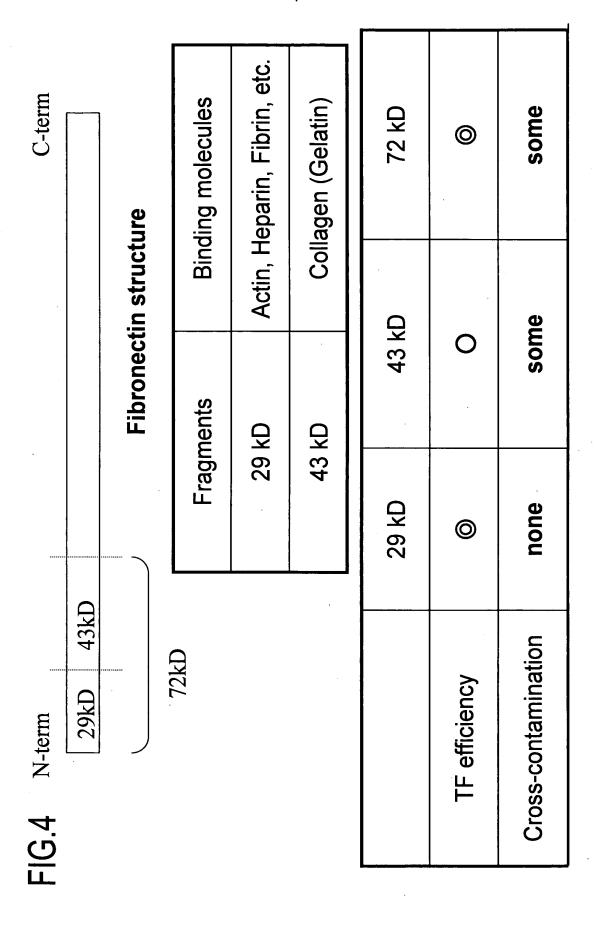


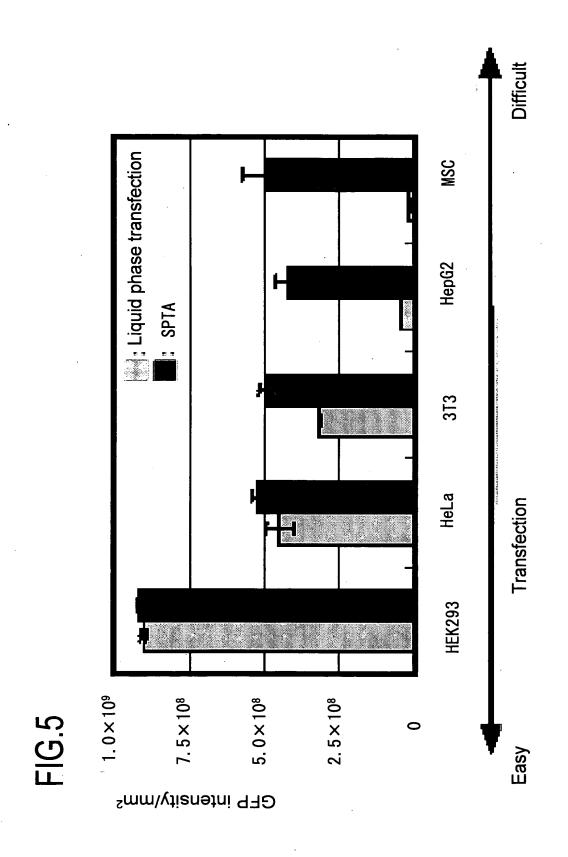
FIG.2

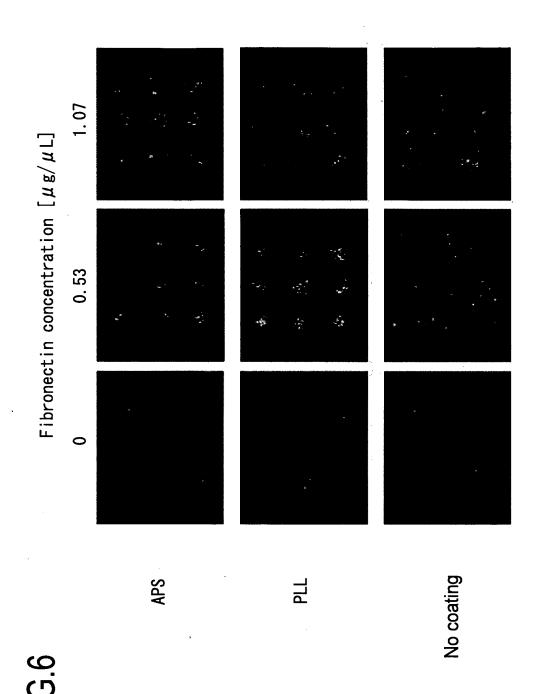


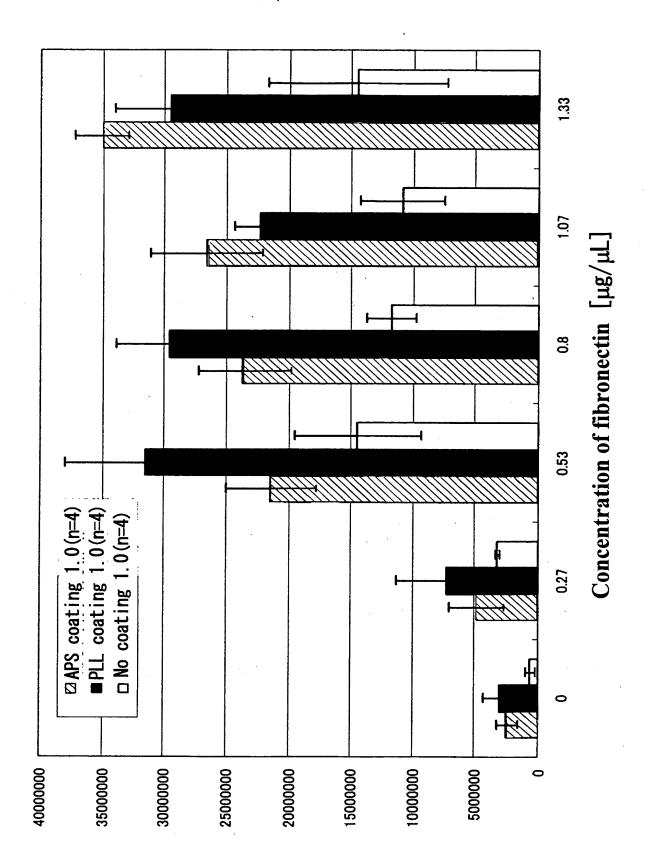






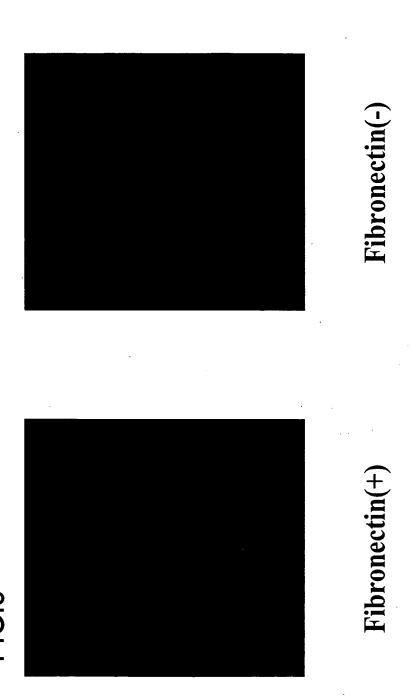


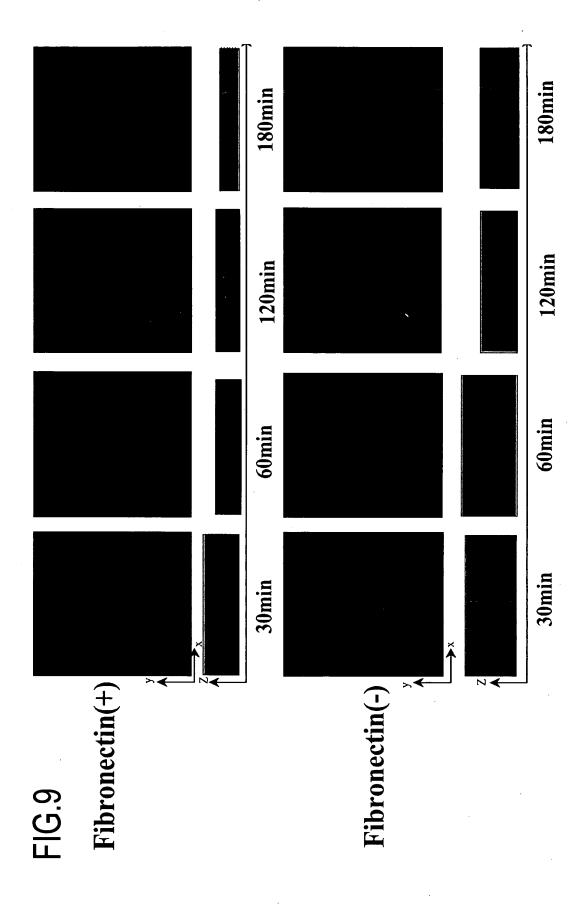




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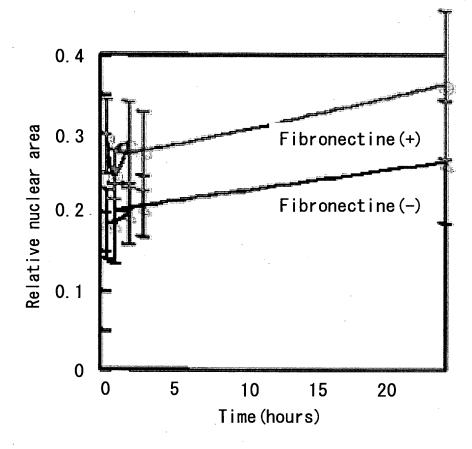


FIG.11

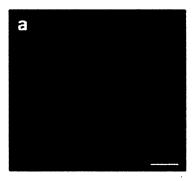
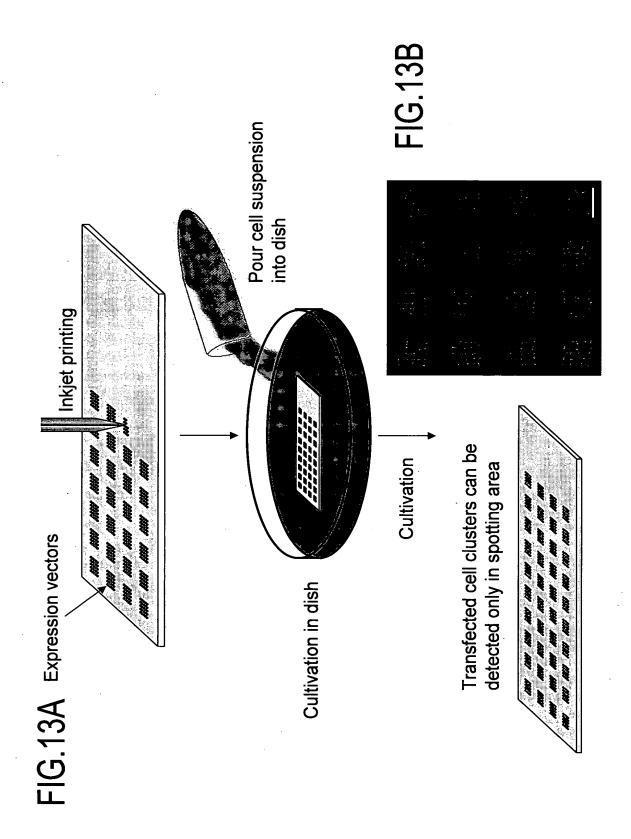
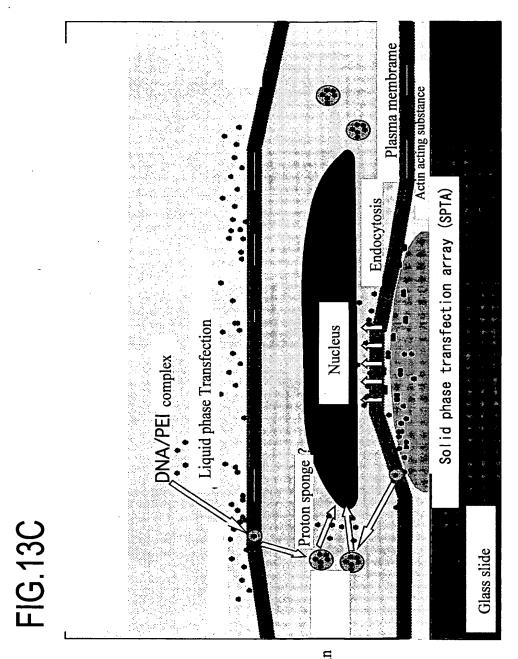


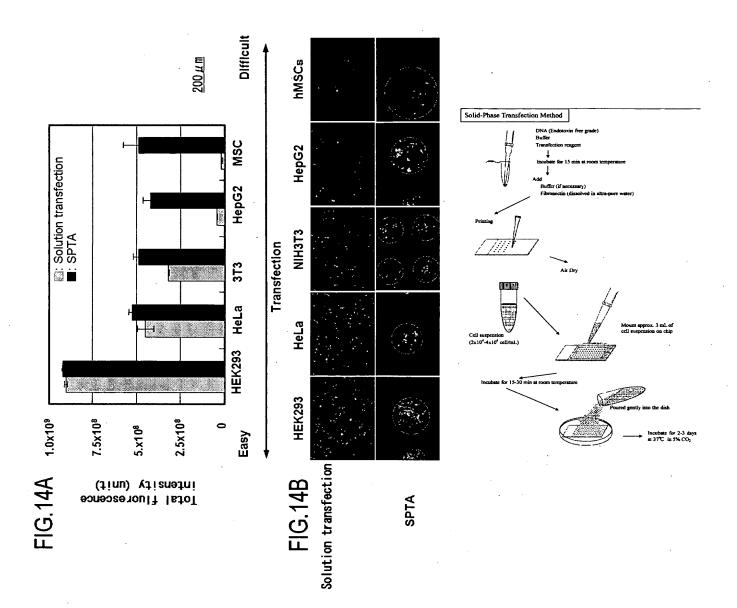
FIG.12







Concentration in lysosome



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FIG.14D

For HEK293		
DMEM (serum free)	9.5	uL
Plasmid DNA (1mg/mL)	1.5	uL
TransFast (1mg/mL)	9.0	uL
DMEM (serum free)	5.0	uL
Fibronectin (4mg/mL)	5.0	uL
Final volume	30.0	υĪ.

For HeLa, NIH3T3-3, HepG2

DMEM (serum free)	14.5	uL
Plasmid DNA (1mg/mL)	1.5	uL
Lipofectamine2000	4.5	uL
DMEM (serum free)	5.0	uL
Fibronectin (4mg/mL)	5.0	uL
Final volume	30.0	uL

For hMSCs

	N/P=5	N/P=10	N/P=20	
DMEM (serum free)	12.75	12.0	10.5	uL
Plasmid DNA (1mg/mL)	1.5	1.5	1.5	uL
JetPEI (x4) conc.	0.75	1.5	3.0	uL
Fibronectin (4mg/mL)	5.0	5.0	5.0	υL
Final volume	20.0	20.0	20.0	uL

Scheme for HEK293

1.5mL micro-tube	
↓ ←DMEM	
↓ ←Plasmid Dl	NA .
mix	Incubate for 2-3 days
↓ ← TransFast	at 37℃ in 5% CO ₂
mix completely	and incubate for 15 min at RT
↓ ← DMEM	
↓ ← Fibronectin	1
mix completely	
1	•
ready to print	

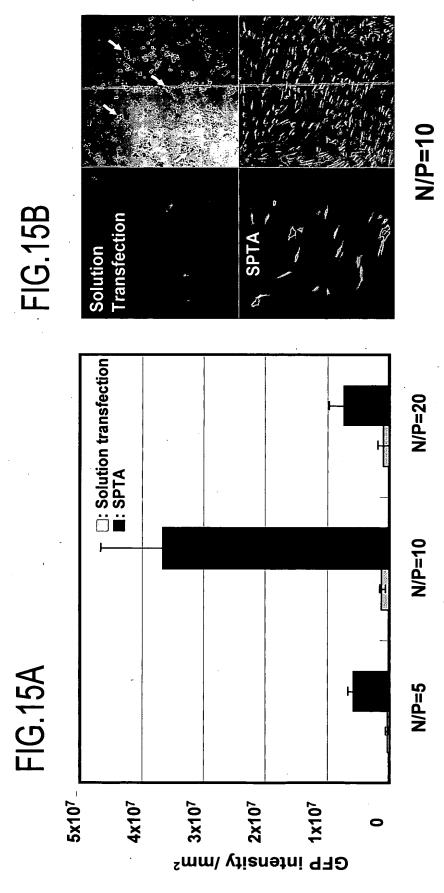
Scheme for HeLa, NIH3T3-3, and HepG2

or rioba, ranis is s, and riopos
1.5mL micro-tube
↓←DMEM
↓ ←Plasmid DNA
mix
↓ ←Lipofectamine2000
mix completely and incubate for 15 min at RT
↓ ←DMEM
↓ ←Fibronectin
mix completely
1
ready to print

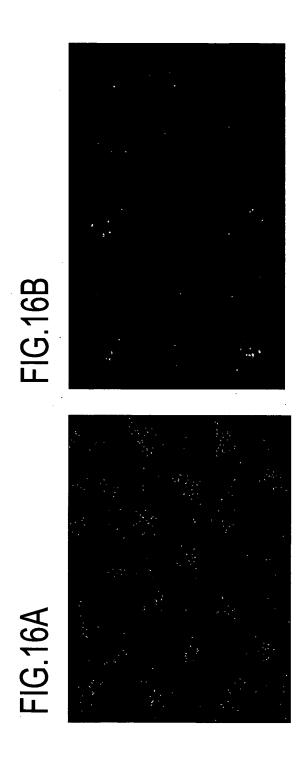
Scheme for hMSCs

ne for niviscs
1.5mL micro-tube
↓ ←DMEM
↓ ←Plasmid DNA
mix
↓ ← jetPEI
mix completely and incubate for 15 min at RT
↓ ←Fibronectin
mix completely
1
ready to print

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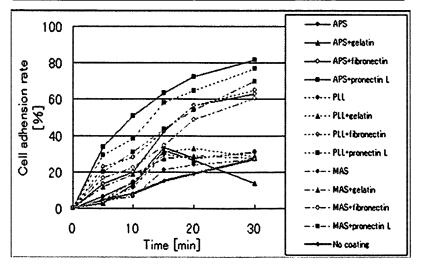
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FIG.16C

Number of adherent co	ells					
Transpor or Bulletone of	Time(min)					
	Ó	5	10	15	20	30
APS	235	220	202	157	170	162
APS+gelatin	212	206	184	145	156	183
APS+fibronectin	229	198	183	132	100	85
APS+pronectin L	257	170	126	94	71	47
						,
PLL	231	221	205	162	168	159
PLL+gelatin	218	208	186	151	146	156
PLL+fibronectin	225	174	162	129	98	79
PLL+pronectin L	214	151	132	90	76	50
1440	001	000	010	100	170	100
MAS	231	222	216	182	176	169
MAS+gelatin	224	198	182	163 143	159	162
MAS+fibronectin	218	182 176	169 152	124	112 101	86 66
MAS+pronectin L	220	1/0	132	124	101	00
No coating	226	216	208	192	183	164
Cell adhension rate (p	roportion of a	herent cel	lls (%))			
	Time(min)			·		
	0	5	10	15	20	30
APS	0	6.382979	14.04255	33.19149	27.65957	31.06383
APS+gelatin	0	2.830189		31.60377	26.41509	13.67925
APS+fibronectin	. 0	13.53712	20.08734		56.33188	62.8821
APS+pronectin L	0	33.85214	50.97276	63.42412	72.37354	81.71206
PLL		4.329004	11.25541	29.87013	27.27273	31.16883
	0	4.529004	14.6789	30.73394	33.02752	
PLL+gelatin						28.44037
PLL+fibronectin	0	22.66667	28 38.31776		56.44444	64.88889
PLL+pronectin L	_ U	29.43925	38.31770]	57.94393	64.48598	76.63551
MAS	0	3.896104	6.493506	21.21212	23.80952	26.83983
MAS+gelatin	0	11.60714		27.23214	29.01786	27.67857
MAS+fibronectin	0	16.51376	22.47706		48.62385	60.55046
MAS+pronectin L	0	20		43.63636	54.09091	70
No coating	0	4.424779	7.964602	15.04425	19.02655	27.43363



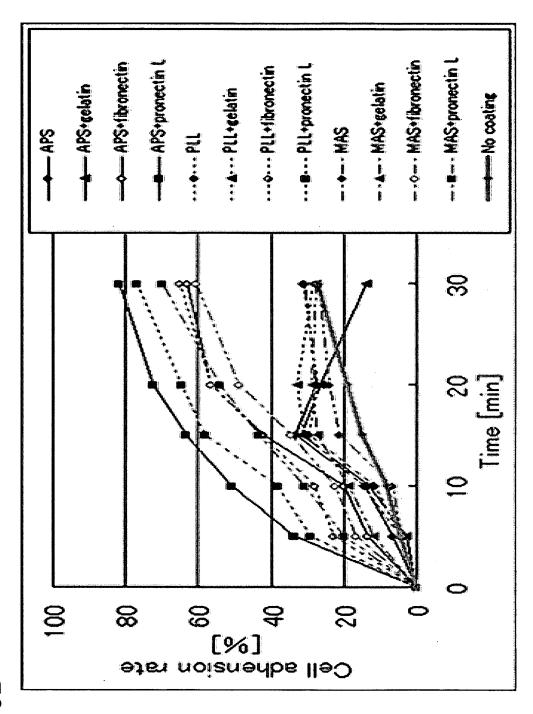


FIG.16D

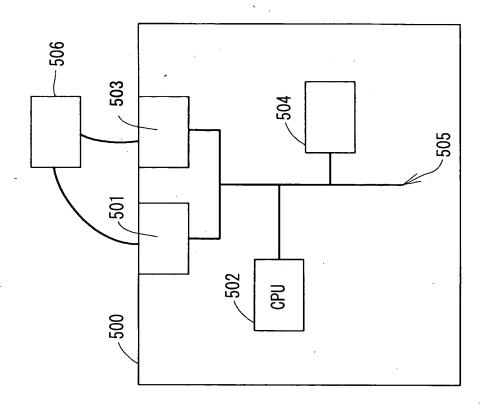


FIG.17

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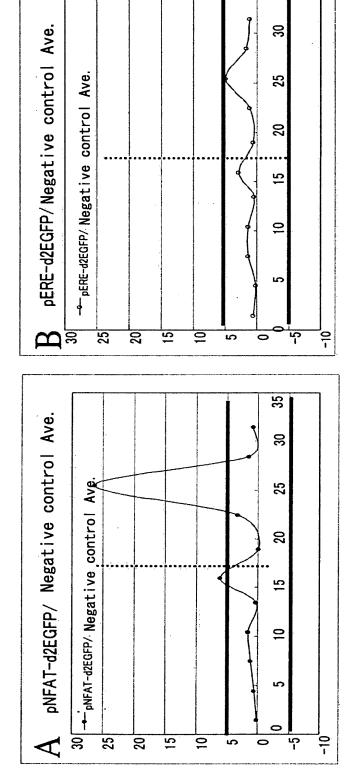
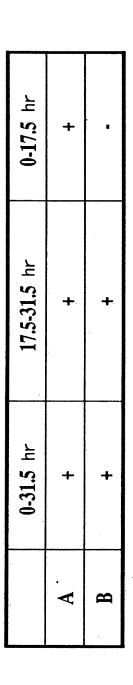
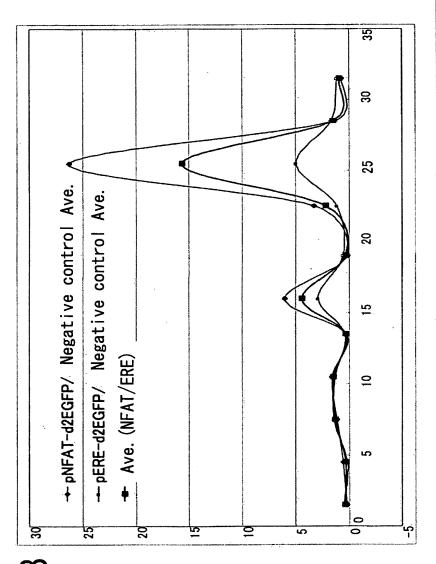


FIG.18A



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	0-31.5 hr	17.5-31.5 hr	0-17.5 hr
NFAT	+	+	+
ERE	+	•	•
NFAT/ERE	+	+	•

FIG.18B

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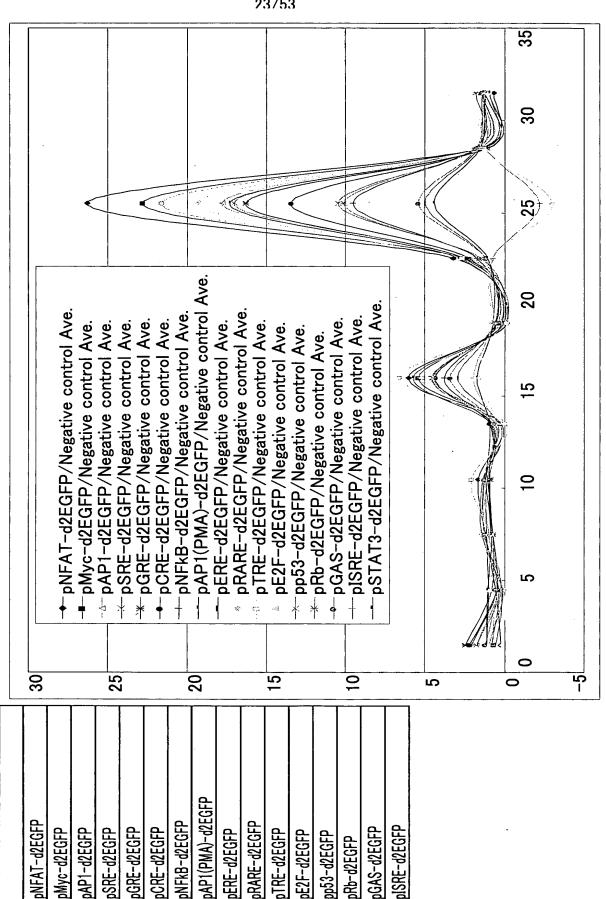


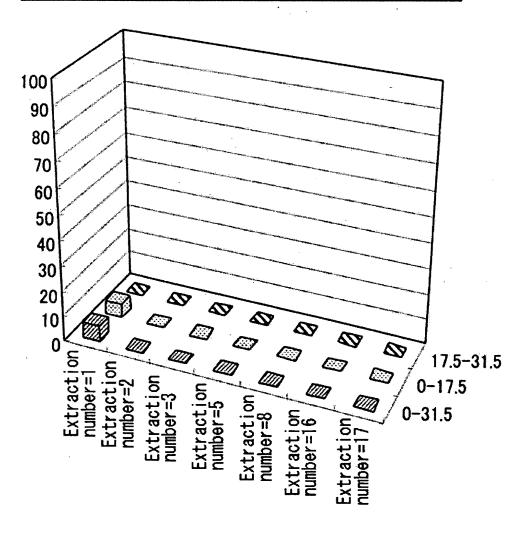
FIG.19

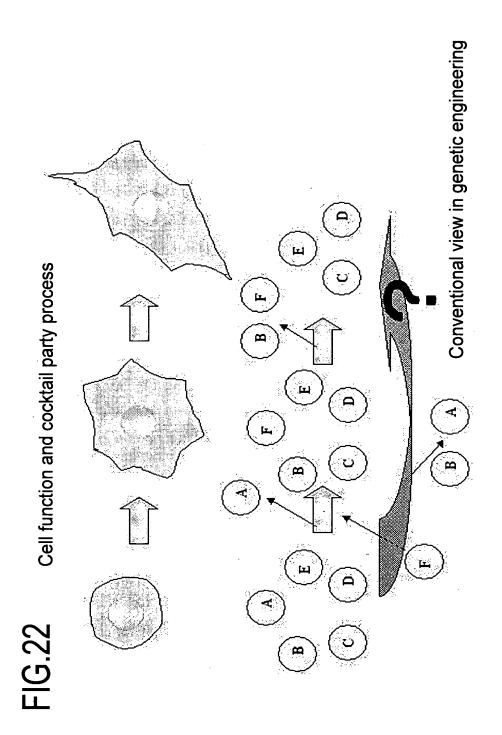
Docket No. 690121.405USPC Inventor(s): Masato Miyake et al. Express Mail No. EV718247128US

	17.5-31.5	82.35294	88.23529	94.11765	94.11765	100	100	100	24/53
	0-17.5	29.41176	41.17647	29.41176	11.76471	5.882353	0	0	
	0-31.5	82.35294	70.58824	88.23529	94.11765	9	100	100	
Day0-1	Induction of differentiation	Extraction number=1	Extraction number=2	Extraction number=3	Extraction number=5	Extraction number=8	Extraction number=16	Extraction number≂17	
TH=5									
		L		E E	001				Extraction number=3 Extraction number=5 Extraction number=8 Extraction number=16 Extraction number=16 Extraction number=16

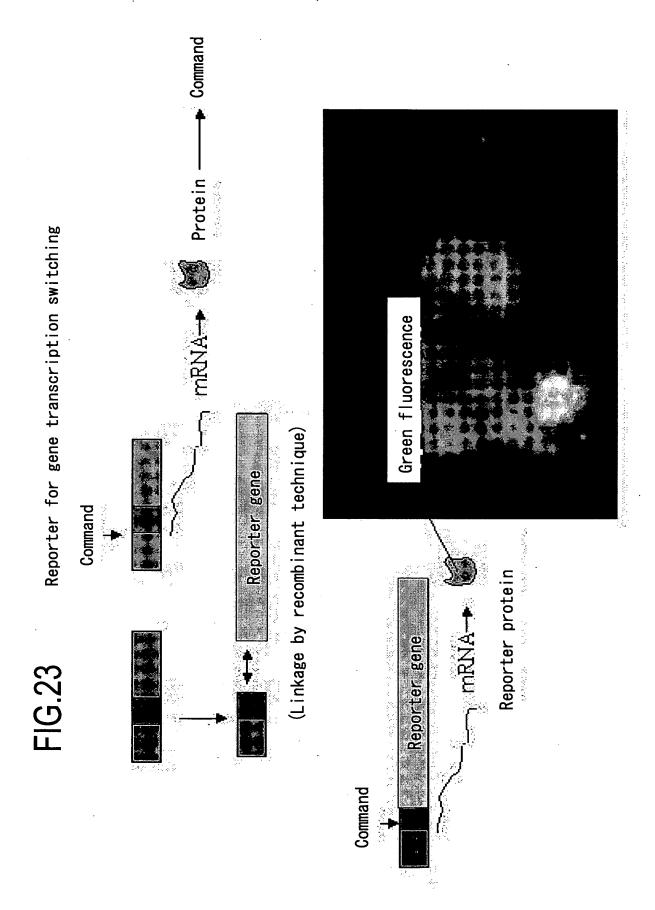
FIG.21

No induction of differentiation	0-31.5	0-17.5	17.5-31.5
Extraction number=1	5.882353	5.882353	C
Extraction number=2	0	. 0	Q
Extraction number=3	0	. 0	C
Extraction number=5	. 0	0	q
Extraction number=8	0	O	C
Extraction number=16	O	0	d
Extraction number=17	0	0	Q





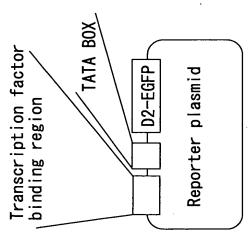
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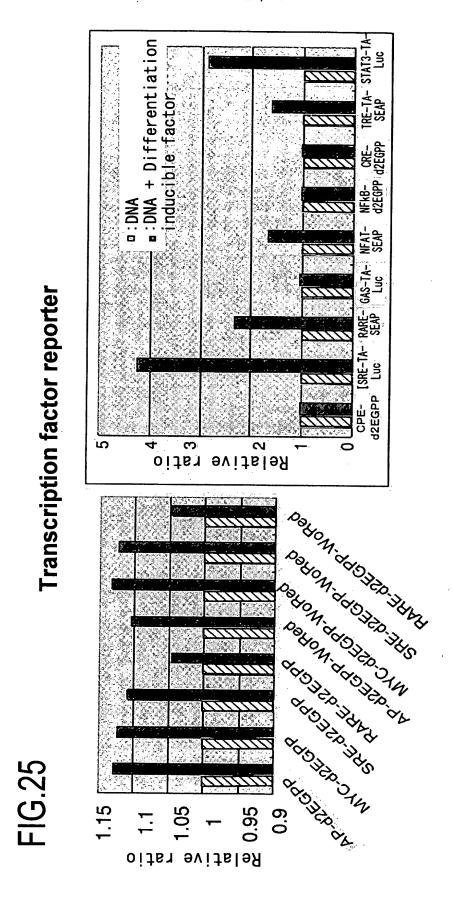
Construction of transcription factor reporter

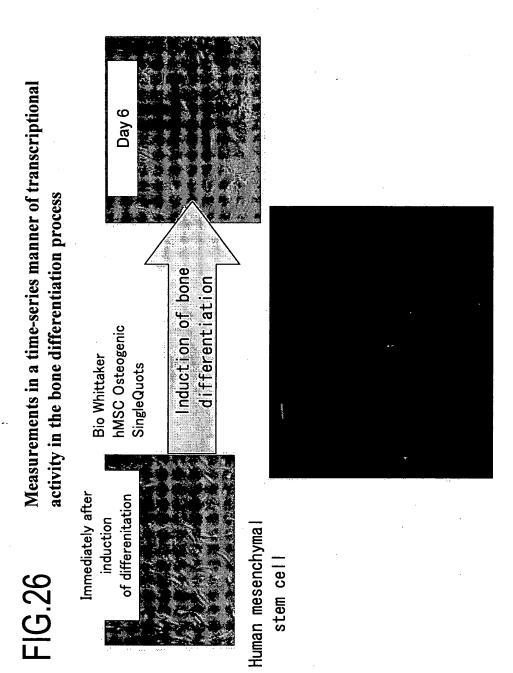
FIG.24

Vector	Pathway	Transcription factor	Cis-acting enhancer element
pNFkB-d2FGFP	IKK/NFkB	NFKB	kВ
pAP1-d2FGFP	SAPKJUK	c—Jun, c-Fos	AP1
pSRF-d2FGFP	MAPK/JNK, MAPK/FRK	FIK-1,STAT, TCF,SRF	SRF
pGRF-d2FGFP	Glicocorticoide (HXP90 mediation)	GR	GRF
pCRF-d2FGFP	PKA/CRFB,JNK/p38 PKA	ATF2/CRFB	CRF
pMpc-TA-d2FGFP, pMYC-,d2FGFP	Cell cycle	c-myc	F-box
pHSF-d2FGFP	HSF	HSF	HSF
pNFAT-d2FGFP	NFAT/Calcineurin/PKC	NFAT	NFAT
pAP1(PMA)-TA-d2FGFP	PKC		AP1(PMA)
pRb-TA-d2FGFP	Cell cycle		Rb
pF2F-TA-d2FGFP	Cell cycle		F2F
pp53-TA-d2FGFP	Cell cycle apoptosis		P53
pGAN-TA-d2FGFP	JAK/STAT	STAT1/STAT1	GAS
pISRF-TA-d2FGFP	JAK/STAT	STAT2/STAT1	ISRF
pSTAT3-TA-d2FGFP	JAK/STAT	STAT3/STAT3	STAT3
pFRF-TA-d2FGFP	Estrogen receptor		FRF
pRARF-TA-d2FGFP	Retinoic acid		RARF
pTRF-TA-d2FGFP	Thyroid receptor		TRF



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TF array culturing chamber

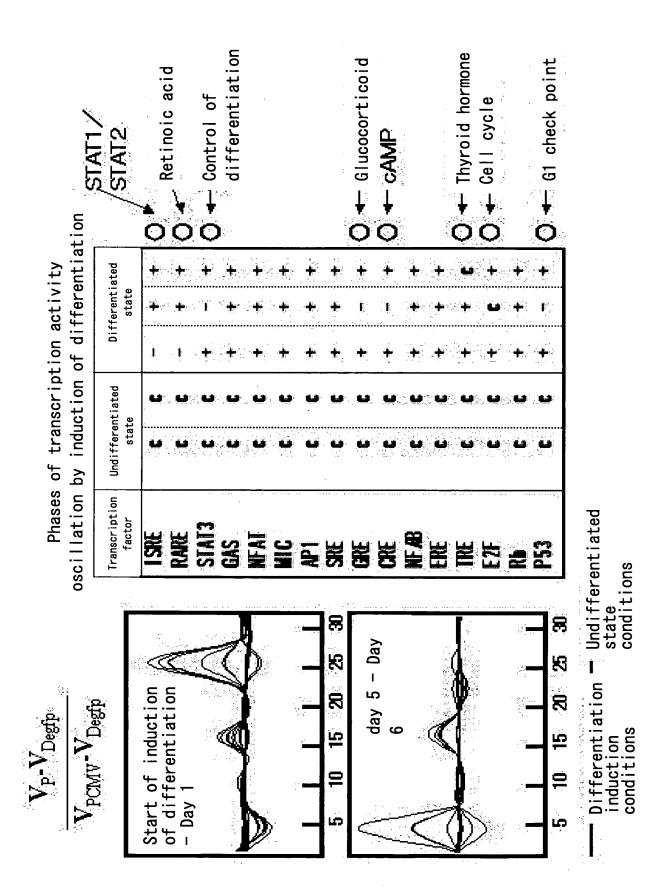
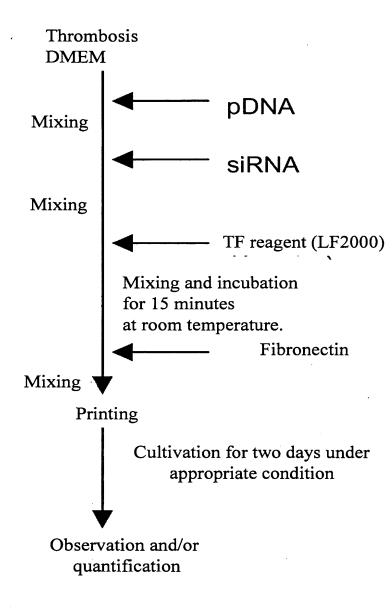
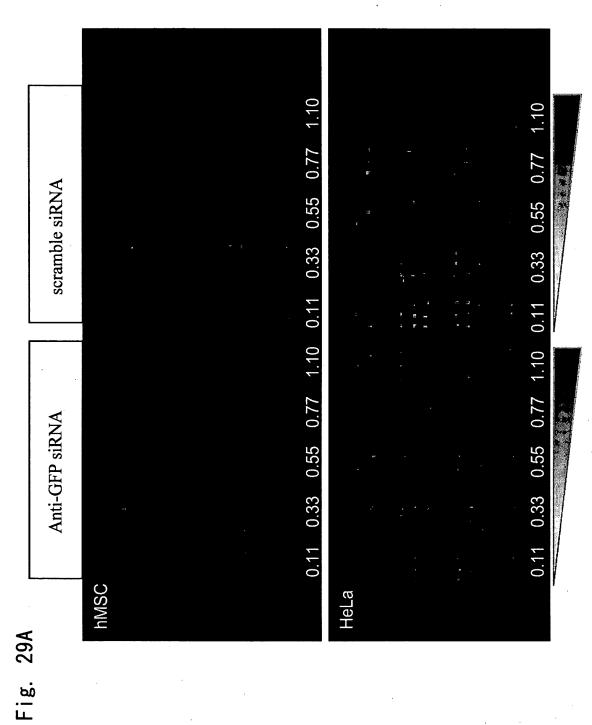


FIG.27

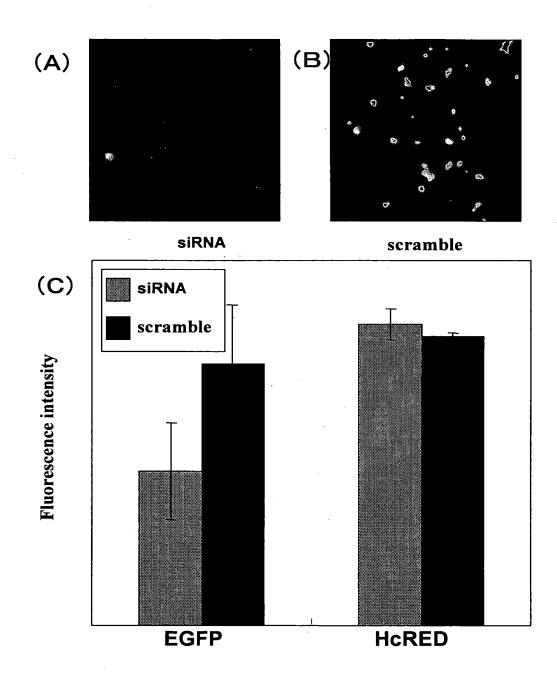
FIG.28



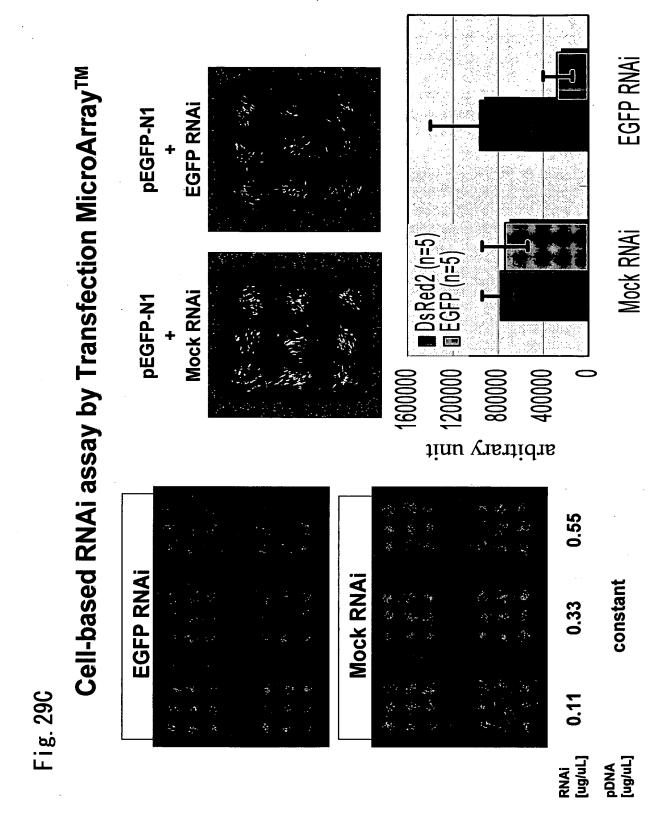


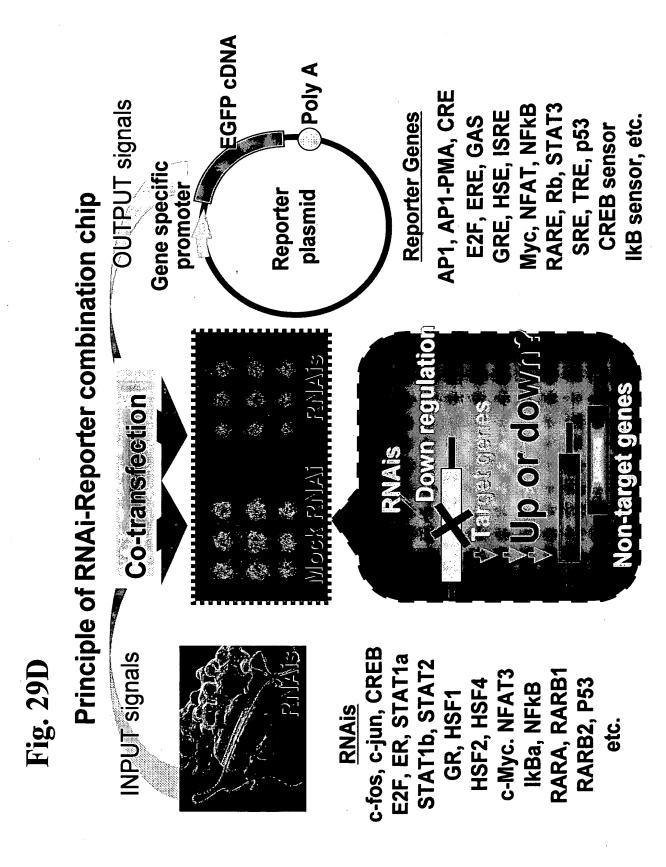
siRNA concentration [μg/μL]

Fig. 29B

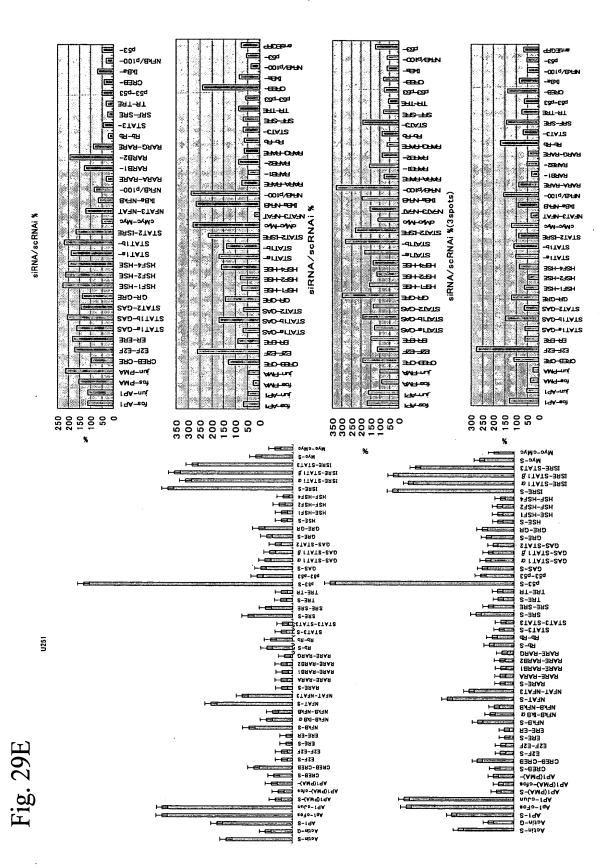


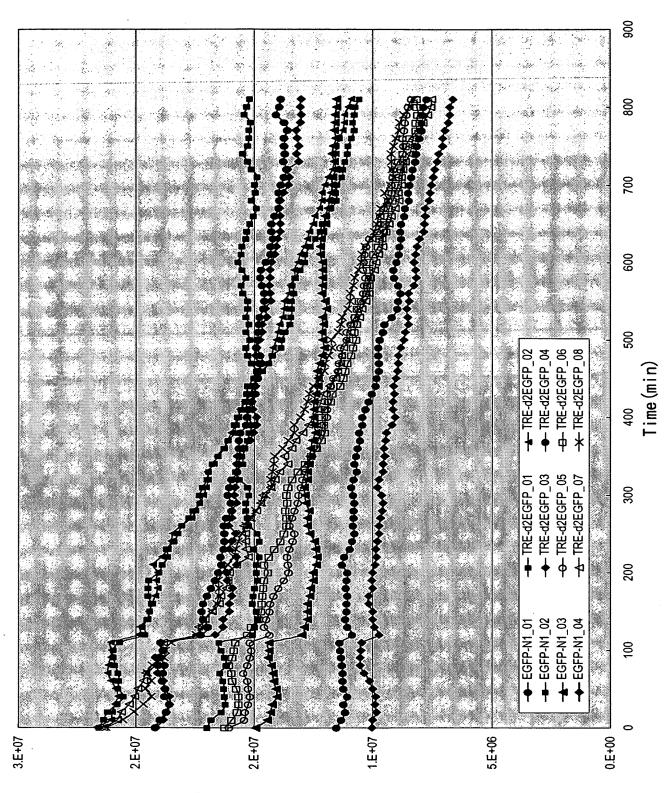
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Total intensity per unit grid

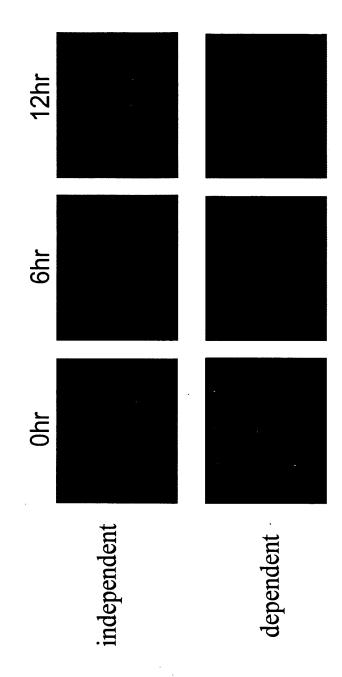
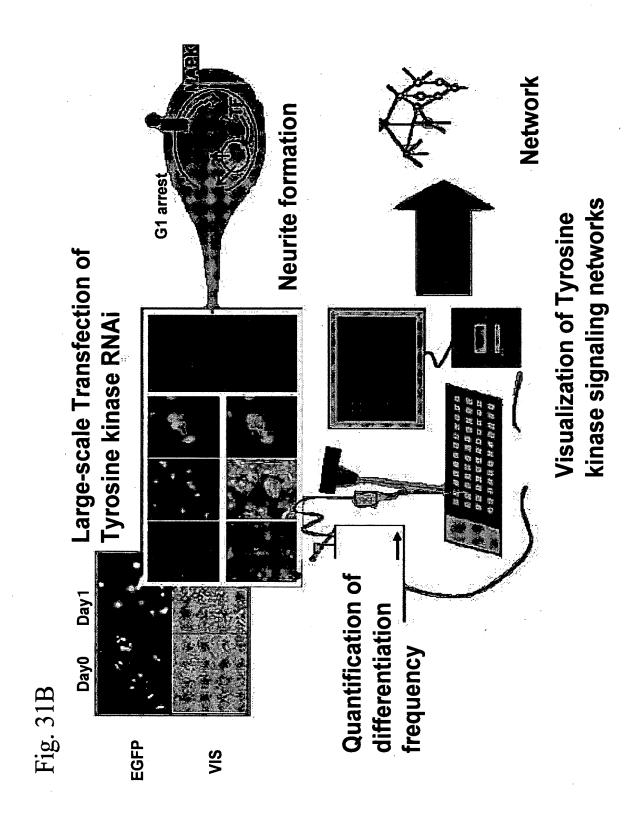
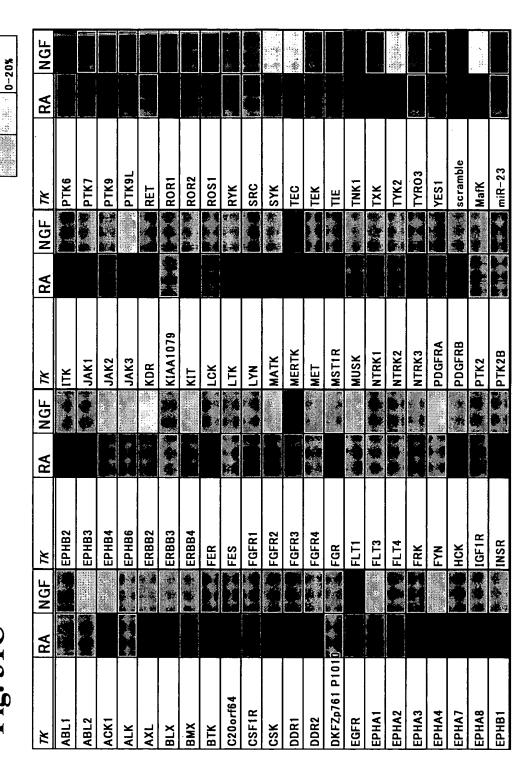


Fig. 31



80-100% 60-80% 40-60% 20-40%

Fig. 31C



(*B-bridge siRNAs against Tyrosine kinases)

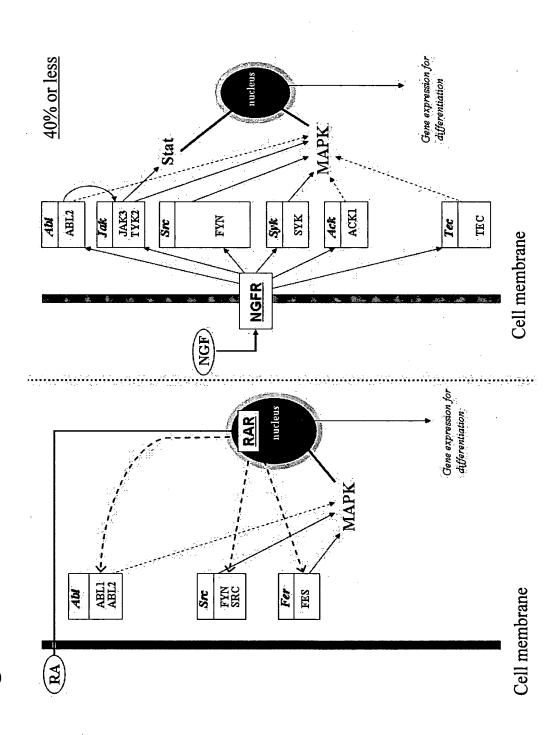


Fig. 311

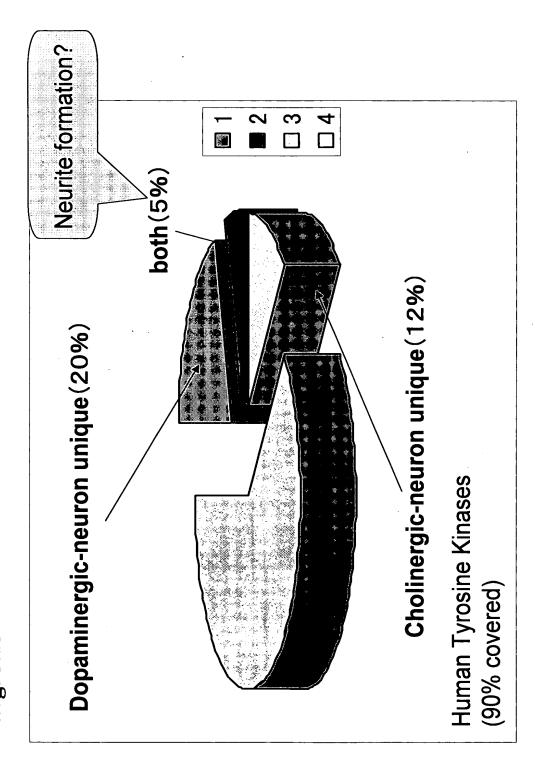


Fig. 31E

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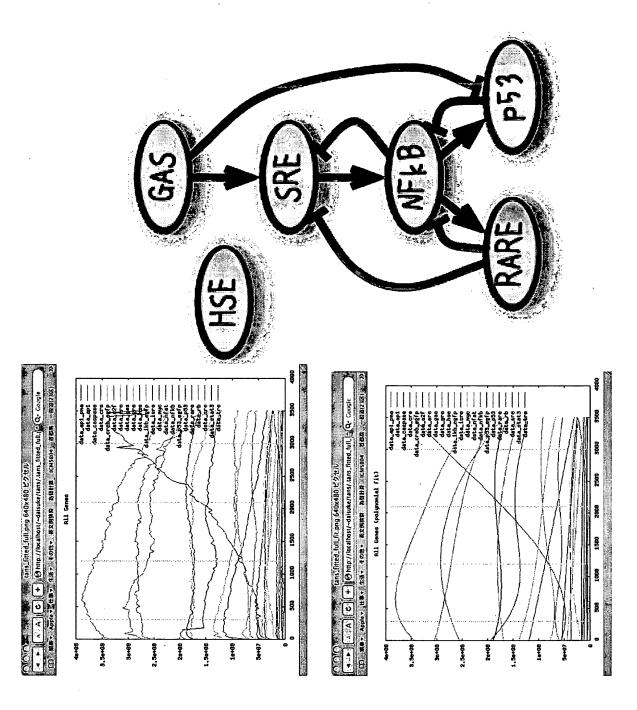


Fig. 31F

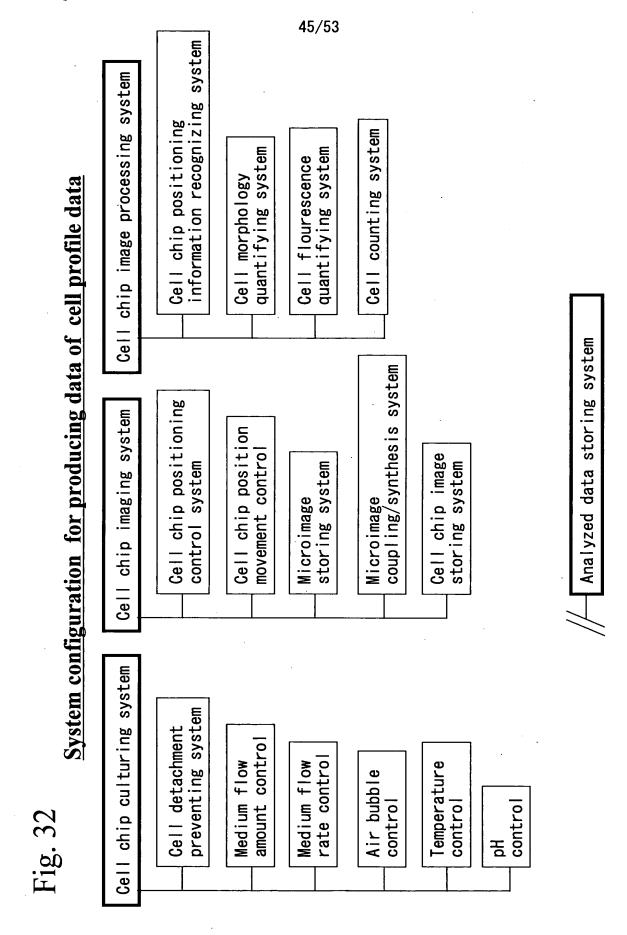
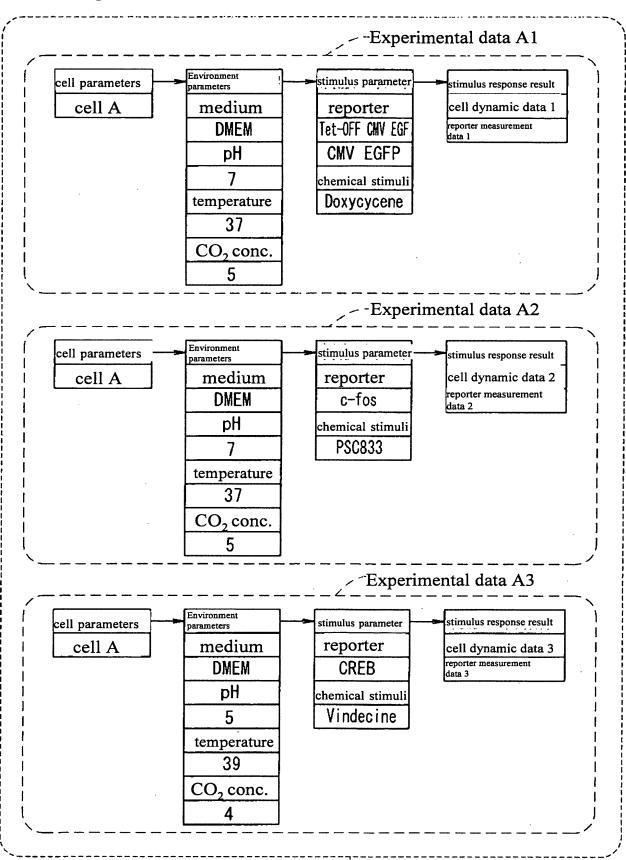


Fig. 33A

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` Digital Cell

Fig. 33B

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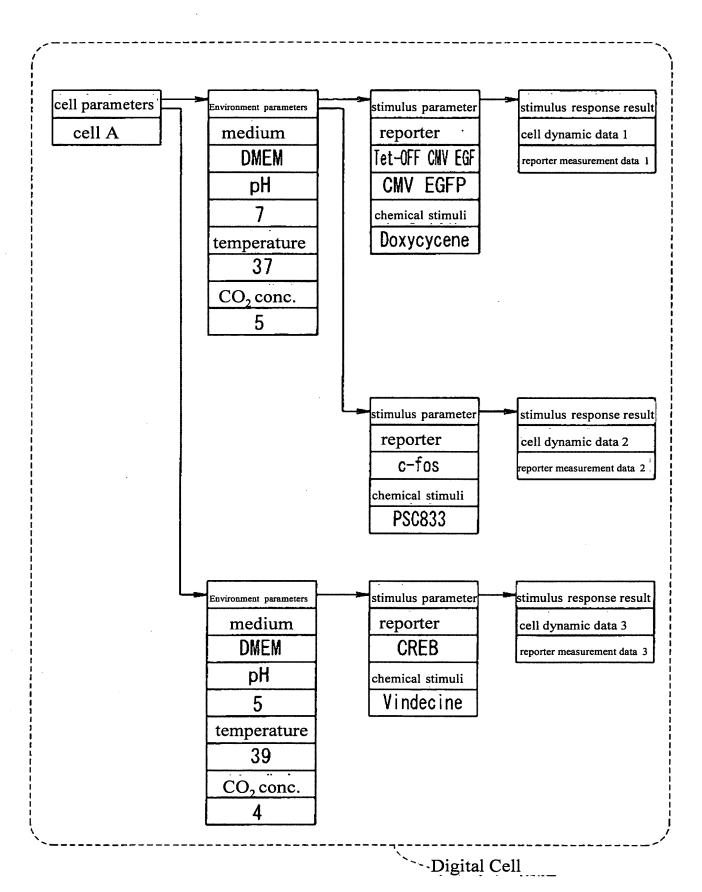


Fig. 34

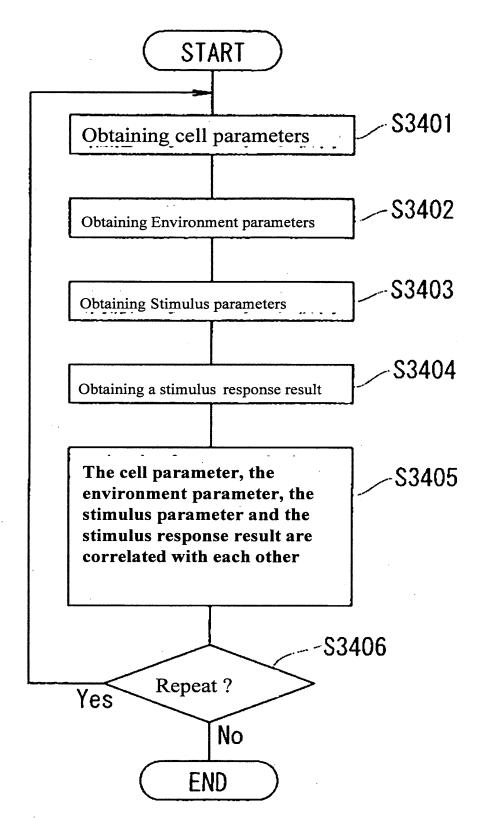


Fig. 35

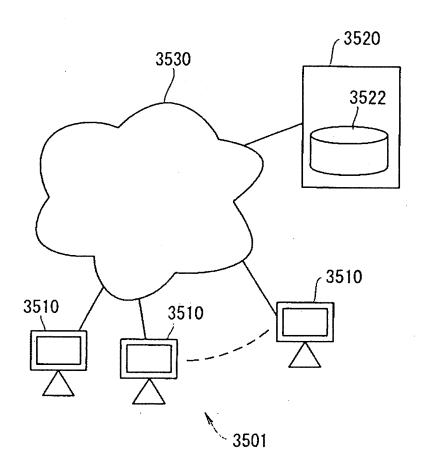
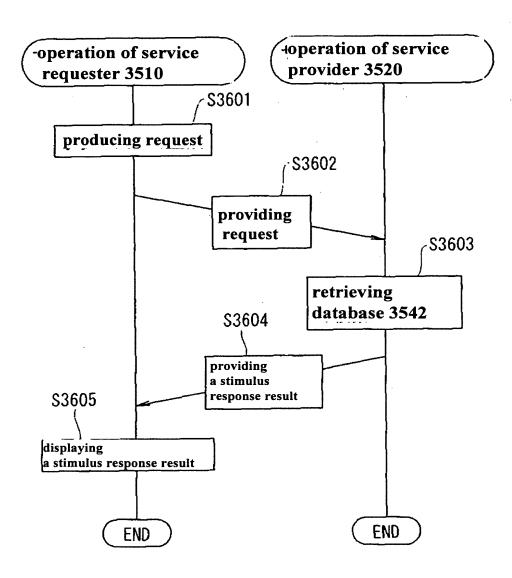


Fig. 36



Input interface for inputting parameters Environment parameters Stimulus parameters Cell parameters

Fig. 37

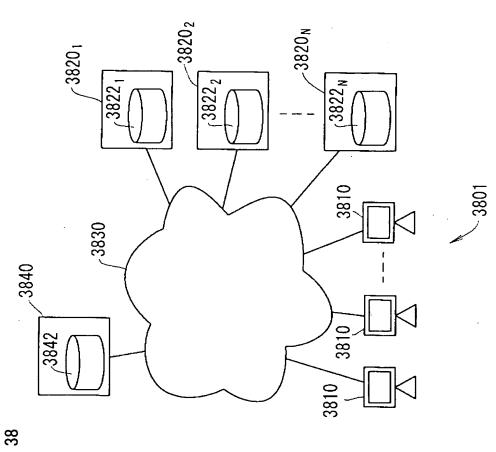
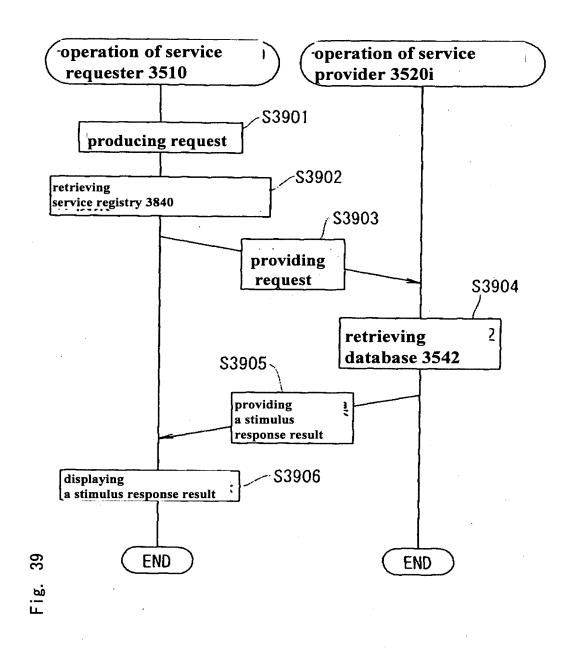


Fig.

4



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